

MODE OPERATOIRE / OPERATING PROCEDURE	VERSION
<b>METHODS OF USING ANNEXIN V FOR FLOW CYTOMETRY</b>	A

### Materials: preparation of positive control

1. Prepare Camptothecin stock solution (Sigma-Aldrich Cat. No. C-9911): 1 mM in DMSO.
2. Jurkat T cells (ATCC TIB-152).

### Procedure

1. Add Camptothecin (final conc. 4-6  $\mu$ M) to  $1 \times 10^6$  Jurkat cells .
2. Incubate the cells for 4-6 hr at 37°C.
3. Proceed with the FITC Annexin V Staining Protocol to measure apoptosis.

### Reagents

1. FITC Annexin V (cat. no. 556420, 556419). Use 5  $\mu$ l per test.
2. Propidium Iodide (PI) (cat. no. 556463). is a convenient, ready-to-use nucleic acid dye. Use 5  $\mu$ l per test.
3. 10X Annexin V Binding Buffer (cat. no. 51-66121E): 0.1 M Hepes/NaOH (pH 7.4), 1.4 M NaCl, 25 mM CaCl<sub>2</sub>. For a 1X working solution, dilute 1 part of the 10X Annexin V Binding Buffer to 9 parts of distilled water.
4. Purified Recombinant Annexin V (cat. no. 554781). Use 5-15  $\mu$ g per test. Researchers are encouraged to titrate the reagent for optimal results.

### Staining

1. Wash cells twice with cold PBS and then resuspend cells in 1X Binding Buffer at a concentration of  $1 \times 10^6$  cells/ml.
2. Transfer 100  $\mu$ l of the solution ( $1 \times 10^5$  cells) to a 5 ml culture tube.
3. Add 5  $\mu$ l of FITC Annexin V and 5  $\mu$ l PI.
4. Gently vortex the cells and incubate for 15 min at RT (25°C) in the dark.
5. Add 400  $\mu$ l of 1X Binding Buffer to each tube. Analyze by flow cytometry within 1 hr.

### Blocking

1. Wash cells twice with cold PBS and then resuspend cells in 1X Binding Buffer at a concentration of  $1 \times 10^6$  cells/ml.
2. Transfer 100  $\mu$ l of the solution ( $1 \times 10^5$  cells) to a 5 ml culture tube.
3. Add 5-15  $\mu$ g of purified recombinant Annexin V. The amount of purified recombinant Annexin V required to saturate binding sites may vary according to cell type and stage of apoptosis. In some cases, investigators may need to reduce the number of cells to  $0.5 \times 10^5$  and still add 5-15  $\mu$ g of recombinant Annexin V to obtain optimal results. Titration is strongly recommended.
4. Gently vortex the cells and incubate for 15 min at room temperature.
5. Add 5  $\mu$ l FITC Annexin V and 5  $\mu$ l PI.
6. Gently vortex the cells and incubate for 15 min at room temperature in the dark.
7. Add 400  $\mu$ l of 1X Binding Buffer to each tube. Analyze by flow cytometry as soon as possible (within 1 hr).

